

Lab 9: Clustering

ACTL3142 and ACTL5110

Questions

Conceptual Questions

1. (ISLR2, Q9.2) * Suppose that we have four observations, for which we compute a dissimilarity matrix, given by

$$\begin{bmatrix} & 0.3 & 0.4 & 0.7 \\ 0.3 & & 0.5 & 0.8 \\ 0.4 & 0.5 & & 0.45 \\ 0.7 & 0.8 & 0.45 & \end{bmatrix}$$

For instance, the dissimilarity between the first and second observations is 0.3, and the dissimilarity between the second and fourth observations is 0.8.

- a. On the basis of this dissimilarity matrix, sketch the dendrogram that results from hierarchically clustering these four observations using complete linkage. Be sure to indicate on the plot the height at which each fusion occurs, as well as the observations corresponding to each leaf in the dendrogram.
- b. Repeat (a), this time using single linkage clustering.
- c. Suppose that we cut the dendrogram obtained in (a) such that two clusters result. Which observations are in each cluster?
- d. Suppose that we cut the dendrogram obtained in (b) such that two clusters result. Which observations are in each cluster?
- e. It is mentioned in the chapter that at each fusion in the dendrogram, the position of the two clusters being fused can be swapped without changing the meaning of the dendrogram. Draw a dendrogram that is equivalent to the dendrogram in (a), for which two or more of the leaves are repositioned, but for which the meaning of the dendrogram is the same.

[Solution](#)

2. (ISLR2, Q9.3) ★ In this problem, you will perform K -means clustering manually, with $K = 2$, on a small example with $n = 6$ observations and $p = 2$ features. The observations are as follows.

Obs	X_1	X_2
1	1	4
2	1	3
3	0	4
4	5	1
5	6	2
6	4	0

- Plot the observations.
- Randomly assign a cluster label to each observation. You can use the `sample()` command in R to do this. Report the cluster labels for each observation.
- Compute the centroid for each cluster.
- Assign each observation to the centroid to which it is closest, in terms of Euclidean distance. Report the cluster labels for each observation.
- Repeat (c) and (d) until the answers obtained stop changing.
- In your plot from (a), color the observations according to the cluster labels obtained.

[Solution](#)

Additional Questions

1. ★ In hierarchical clustering, two dissimilarity measures have been mentioned: Euclidean and correlation.

Consider two vectors $\mathbf{x} = (x_1, x_2, \dots, x_n)^T$ and $\mathbf{y} = (y_1, y_2, \dots, y_n)^T$

Their Euclidean dissimilarity is: $(\sum_{i=1}^n (x_i - y_i)^2)^{1/2}$

For the correlation dissimilarity, we treat \mathbf{x} values as if they are from the same distribution X and \mathbf{y} values as if they are from the same distribution Y . Hence, their correlation dissimilarity is $1 - \text{Corr}(X, Y)$

- Show that, in the case where $\mathbb{E}[X] = \mathbb{E}[Y] = 0$ and $\sigma_X = \sigma_Y = 1$, correlation dissimilarity and Euclidean dissimilarity give the same results. (Use the population variance instead of the sample variance)

- b. Provide a geometric interpretation of the correlation dissimilarity and the Euclidean dissimilarity. When would each be appropriate?

Solution

Applied Questions

1. (ISLR2, Q9.9) ★ Consider the `USArrests` data. We will now perform hierarchical clustering on the states.
 - a. Using hierarchical clustering with complete linkage and Euclidean distance, cluster the states.
 - b. Cut the dendrogram at a height that results in three distinct clusters. Which states belong to which clusters?
 - c. Hierarchically cluster the states using complete linkage and Euclidean distance, *after scaling the variables to have standard deviation one*.
 - d. What effect does scaling the variables have on the hierarchical clustering obtained? In your opinion, should the variables be scaled before the inter-observation dissimilarities are computed? Provide a justification for your answer.

Solution

2. (ISLR2, Q9.13) On the book website, www.statlearning.com, there is a gene expression data set (`Ch12Ex13.csv`) that consists of 40 tissue samples with measurements on 1,000 genes. The first 20 samples are from healthy patients, while the second 20 are from a diseased group.
 - a. Load in the data using `read.csv()`. You will need to select `header = F`.
 - b. Apply hierarchical clustering to the samples using correlationbased distance, and plot the dendrogram. Do the genes separate the samples into the two groups? Do your results depend on the type of linkage used?

Solution

Solutions

Conceptual Questions

1.
 - a. See Figure 1
 - b. See Figure 2

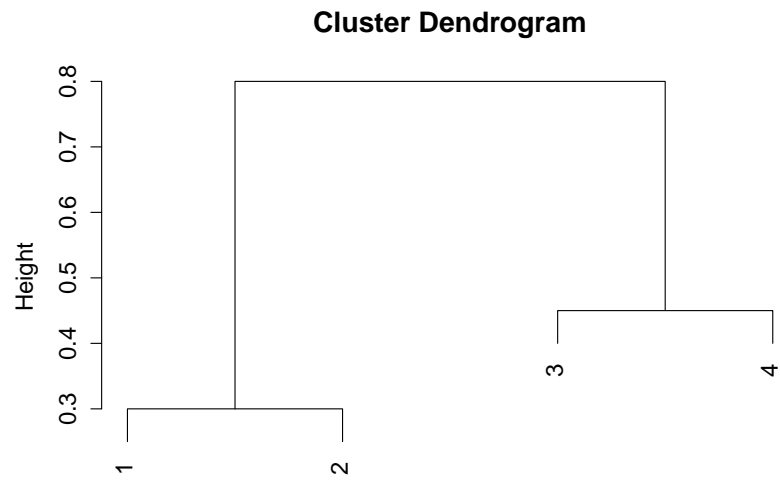


Figure 1: The dendrogram from hierarchically clustering the four observations using complete linkage.

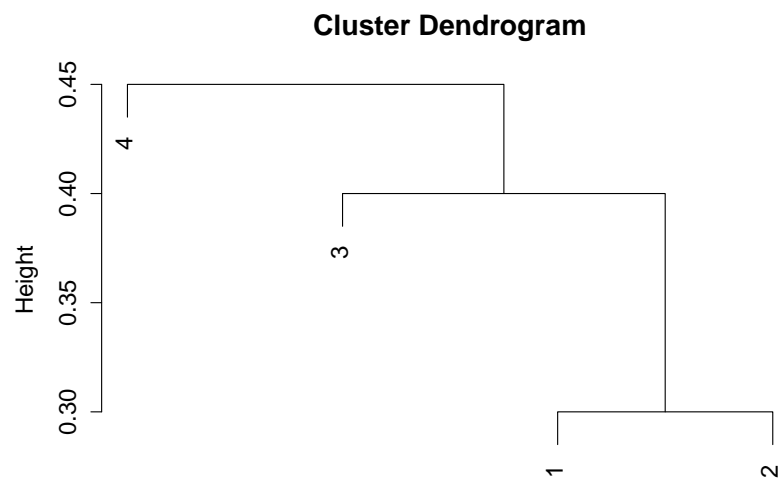


Figure 2: The dendrogram from hierarchically clustering the four observations using single linkage.

- c. (1,2), (3,4)
- d. (1, 2, 3), (4)
- e. See Figure 3

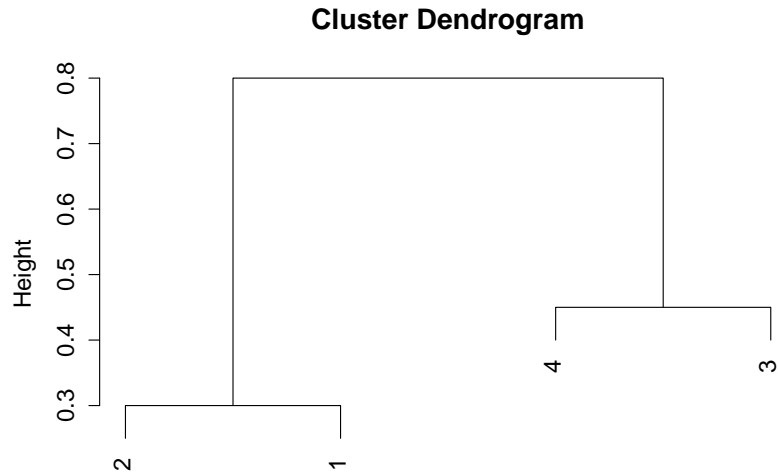


Figure 3: The dendrogram equivalent to Figure 1, for which two or more of the leaves are repositioned, but for which the meaning of the dendrogram is the same.

- 2. a. See Figure 4
- b. See Table 2.

Table 2: Randomly assigned cluster labels to each observation.

Obs.	X_1	X_2	Label
1	1	4	1
2	1	3	1
3	0	4	2
4	5	1	2
5	6	2	1
6	4	0	2

- c. **Label 1** $X_1 = 2.67, X_2 = 3.00$
Label 2 $X_1 = 3.00, X_2 = 1.67$

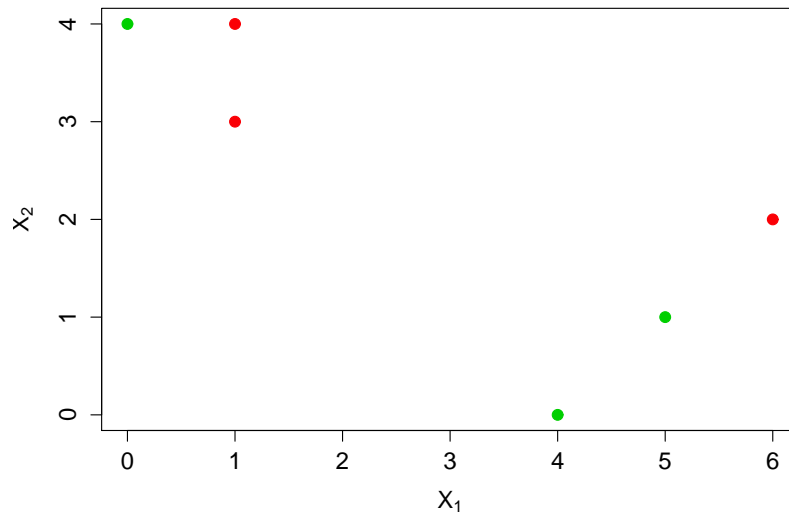


Figure 4: The sample with $n = 6$ observations and $p = 2$ features.

d. See Table 3.

Table 3: Cluster labels to each observation after one iteration.

Obs.	X_1	X_2	Label
1	1	4	1
2	1	3	1
3	0	4	1
4	5	1	2
5	6	2	2
6	4	0	2

e. See Figure 5

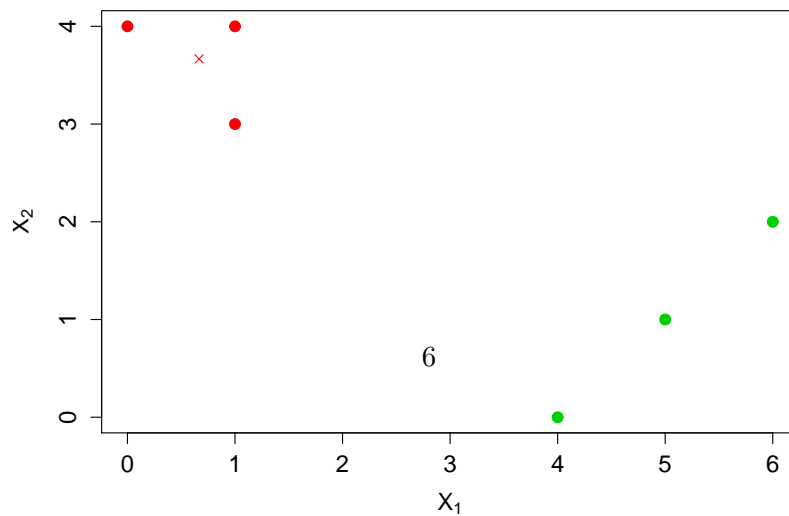


Figure 5: The final result of the K -means clustering.

Noting that $1 = \sigma_X^2 = \mathbb{E}(X^2) - \mathbb{E}(X)^2 = \mathbb{E}(X^2) = \frac{1}{n} \sum_{i=1}^n x_i^2$.
 Now breaking down the Correlation dissimilarity:

$$\begin{aligned} 1 - \text{Corr}(X, Y) &= 1 - \frac{\mathbb{E}((X - \mathbb{E}(X))(Y - \mathbb{E}(Y)))}{\sigma_X \sigma_Y} \\ &= 1 - \mathbb{E}(XY) \\ &= 1 - \frac{1}{n} \sum_{i=1}^n x_i y_i \end{aligned}$$

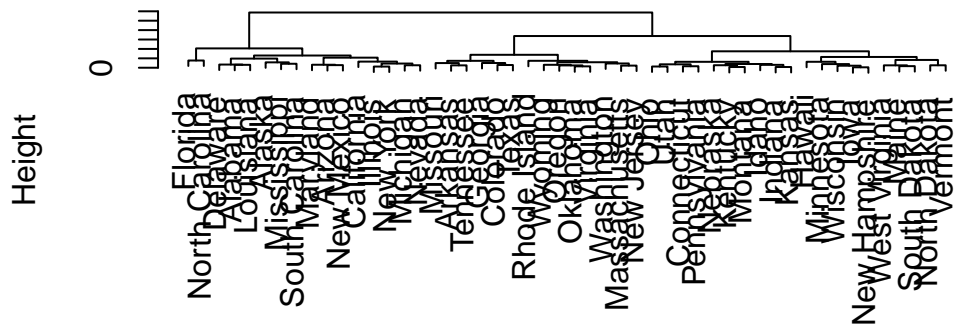
This is directly proportional to the Euclidean dissimilarity, and thus will give the same results (provided the method is linear in the way it summarises).

- b. Euclidean dissimilarity looks at how close the points are to each other in the parameter space, so if the data is clustered and related data tends to be of the same order of magnitude, this is appropriate. Correlation dissimilarity looks at how well the points fit on the same line. This is more appropriate if similar changes in features indicate similarity, despite the values being rather different.

Applied Questions

1. a. `USArrests.hist <- hclust(dist(USArrests), method = "complete")`
`plot(USArrests.hist)`

Cluster Dendrogram



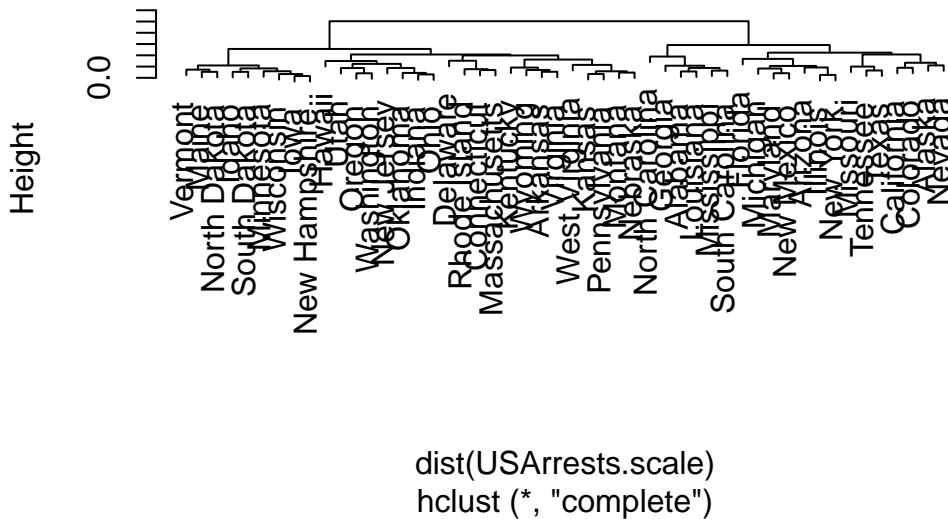
`dist(USArrests)`
`hclust (*, "complete")`

- b. `cutree(USArrests.hist, 3)`

Alabama	Alaska	Arizona	Arkansas	California
1	1	1	2	1
Colorado	Connecticut	Delaware	Florida	Georgia
2	3	1	1	2
Hawaii	Idaho	Illinois	Indiana	Iowa
3	3	1	3	3
Kansas	Kentucky	Louisiana	Maine	Maryland
3	3	1	3	1
Massachusetts	Michigan	Minnesota	Mississippi	Missouri
2	1	3	1	2
Montana	Nebraska	Nevada	New Hampshire	New Jersey
3	3	1	3	2
New Mexico	New York	North Carolina	North Dakota	Ohio
1	1	1	3	3
Oklahoma	Oregon	Pennsylvania	Rhode Island	South Carolina
2	2	3	2	1
South Dakota	Tennessee	Texas	Utah	Vermont
3	2	2	3	3
Virginia	Washington	West Virginia	Wisconsin	Wyoming
2	2	3	3	2

```
c. USArrests.scale <- scale(USArrests, center = FALSE, scale = TRUE)
   USArrests.scale.hist <- hclust(dist(USArrests.scale))
   plot(USArrests.scale.hist)
```

Cluster Dendrogram



d. Scaling reduces the effect of variables which have a significantly higher order of

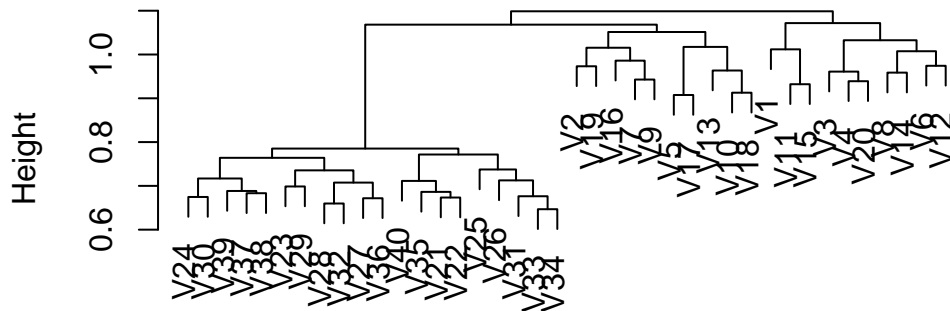
magnitude, especially since a Euclidean distance is used. This will cause some of the values to shuffle categories around the second-third level or lower, but the overall high-level structure remains about the same. Scaling should be used especially when a distance metric is used, since it ensures the values are comparable and have the same (null) unit.

2. a.

```
# Note that you will have to download the csv file into your working  
# directory  
myData <- read.csv("Ch12Ex13.csv", header = FALSE)
```
- b.

```
gene.cluster <- hclust(as.dist(1 - cor(myData)), method = "complete")  
plot(gene.cluster)
```

Cluster Dendrogram



```
as.dist(1 - cor(myData))  
hclust (*, "complete")
```

We note that the splits are very clear for complete linkage. Try some of the other methods to see what they produce.